

Homing Peptides

Screening of a Combinatorial Homing Peptide Library for Selective Cellular Delivery**

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Selective delivery of cargos into specific cell types has the capacity to ensure optimal distribution of therapeutic entities into diseased cells while limiting possible adverse, off-target effects.^[1] One possible route whereby this might be achieved is through receptor-mediated endocytosis, which provides opportunities for targeted delivery where specific receptors are (over)expressed.^[1] Such a strategy has identified the first-generation tumor homing peptides RGD (Arg-Gly-Asp) and NGR (Asn-Gly-Arg), which recognize the α_v integrins and aminopeptidase N, respectively, which are upregulated during tumor progression. These peptides have entered clinical trials.^[2]

Cell-penetrating peptides typically have a high abundance of arginine or lysine residues, or alternating charged and hydrophobic amino acid residues,^[3] with perhaps the most highlighted being the peptide to the transacting activator of transcription (TAT; GRKKRQRRR). TAT has been shown to promote cellular delivery or uptake of conjugated proteins, phage, liposomes, small molecules, and nanoparticles,^[4] and the peptide is able to traverse almost all tissues, including the brain.^[1,3,5] Cell-penetrating peptides are readily synthesized, typically have low immunogenicity,^[5e] and offer numerous opportunities for the delivery of a variety of cargos.^[5d,e]

A chimeric peptide comprising a tumor-homing-peptide section and a cell-penetrating-peptide section has been reported to deliver cargos efficiently inside cells.^[6] A method that would allow the identification of peptides to optimize both the delivery and tumor penetration of existing cancer drugs in a cell-selective manner would be highly desirable. Such an approach would tie in with the desire to generate ligands with affinity for tissue-specific markers, in

essence a “zip code” system for all cell types.^[2f,h,i] Herein, we report a strategy for the high-throughput screening of a peptide nucleic acid (PNA)-encoded peptide library to allow the identification of versatile cell-penetrating homing peptides (Figure 1).

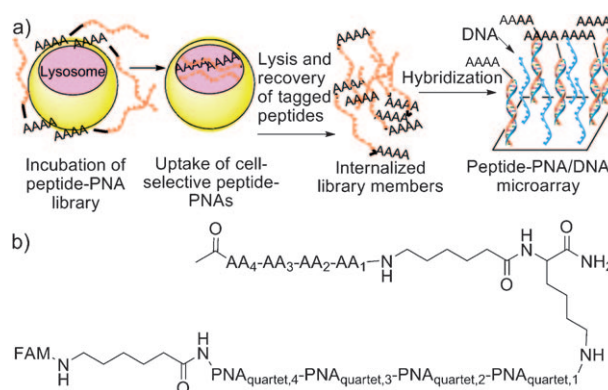


Figure 1. a) The strategy to identify cell-selective penetrating peptides. An encoded 1296-member peptide library was incubated with cells, and any cell-surface-bound peptides were released with trypsin. Cells were lysed, and intracellular PNA was extracted and hybridized onto a 44K DNA microarray. b) General structure of the PNA-encoded 1296-member peptide library (Library 1). Each PNA quartet was designed to have a maximum of 50% similarity to each other quartet and a maximum purine content of 50%; all had uniform melting temperatures and did not include palindrome sequences or polythymine moieties.

A 1296-member carboxyfluorescein (FAM)-labeled, PNA-encoded,^[6,7] tetrapeptide library (Library 1) was designed and synthesized using split and mix methods (see Figure 1 and Scheme S1).^[8] Six amino acids were used, each encoded by a PNA quartet: Pro (turn, AAAC), Glu (acidic, ATCT), Leu (hydrophobic, TACA), Lys (basic, TCAT), and Tyr (aromatic, ACAA) to represent the overall classes of the natural amino acids as well as an *N*-alkyl glycine lysine-like peptoid monomer^[9] (*N*-aminohexylaminoacetic acid (Llp), basic, TTAC; see Scheme S1 in the Supporting Information). This monomer was included because its tetramer is a known highly efficient cellular delivery vehicle that resists enzymatic degradation.^[10]

For library screening and hybridization, cervix epitheloid carcinoma (HeLa), erythroleukemic (K562), embryonic kidney cancer (HEK293T), neuroblastoma (SH-SY5Y), and amelanotic melanoma (ARN8) cells and embryonic stem cells (E14) as well as human primary lymphocytes, monocytes, and neutrophils were incubated with Library 1 (100 μ M; corre-

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sponding to 77 nm of each library member). After incubation, washing, and treatment with trypsin, the cells were lysed, and the library members were extracted and purified by filter centrifugation (between 3000 and 10000 Da) and hybridized onto custom-designed microarrays (44K × 4 Agilent). Microarray scanning and data analysis (BlueFuse, BlueGenome) was used to extract the intensity of the FAM label, thereby giving the relative amount of PNA hybridized to each spot and the identity of the peptide. A generic consensus sequence (Glu-Llp-Glu-Glu) for all cell types was identified by analysis of scatter plots of microarray intensities versus the variable monomers for positions AA₁ to AA₄, a surprise in view of the typically basic residues traditionally observed (see Figure 2 and Figure S1 in the Supporting Information).

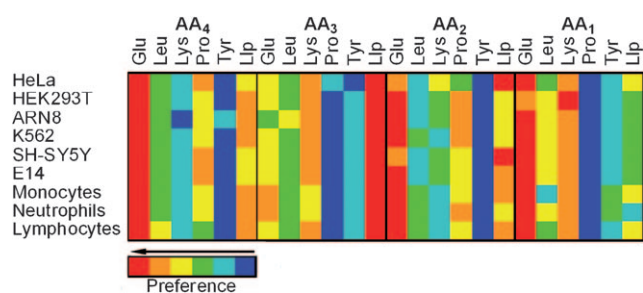


Figure 2. DNA microarray analysis of the extracted members of the peptide–PNA Library 1, which identifies a generic consensus sequence of highly cell-penetrating peptides. Each microarray consisted of four subarrays of 44 000 features each, with 33 replicates of each oligonucleotide complementary to each member of the library as well as 1232 noncoding negative controls. The heat map shows the cell-penetrating peptide preferences extracted from the scatter plots.

To verify and quantitatively compare the cellular delivery abilities, the consensus sequence (FAM-Ahx-Glu-Llp-Glu-Glu-NH₂; the Ahx monomer is 6-aminohexanoic acid) and the positive control (FAM-Ahx-tetraLlp-NH₂) were synthesized^[8a,10a] and incubated with the cells and analyzed by flow cytometry. The identified consensus sequence FAM-Ahx-Glu-Llp-Glu-Glu-NH₂ had uptake levels similar to or greater than FAM-Ahx-tetraLlp-NH₂ in K562, SH-SY5Y, HEK293T, HeLa, and ARN8 cells, thus indicating that this novel highly anionic delivery agent could in some cases result in better delivery than the known delivery agent (Figure 3).

The identification of a generic delivery sequence was useful, but a far more powerful outcome would be the identification of cell-specific peptides. To identify cell-selective peptides, the data for the nine cell types were clustered using Enthought Python and a Euclidean distance method (see the Supporting Information, including Figure S2). This procedure resulted in a dendrogram and heat diagram for each cell type (Supporting Information, Figures S2 and S3), from which cell-selective delivery peptides were derived (Table 1).

To verify and quantitatively compare the cellular delivery abilities of the sequences identified by clustering analysis, the peptides were synthesized with a fluorescent tag: FAM-Ahx-AA₄-AA₃-AA₂-AA₁-NH₂.^[8a,10a] All the cell types were incubated with the nine delivery agents and analyzed by flow

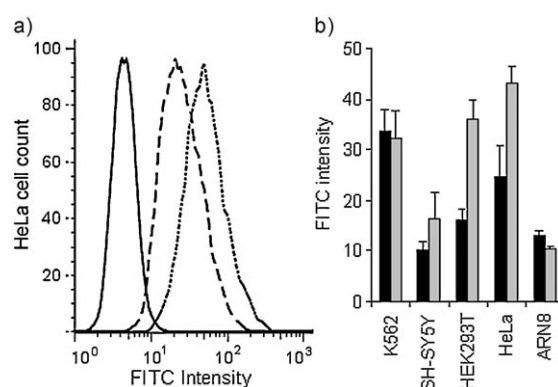


Figure 3. Flow cytometry analysis of the peptides FAM-Ahx-Glu-Llp-Glu-Glu-NH₂ and FAM-Ahx-tetraLlp-NH₂ using fluorescein isothiocyanate (FITC) and phycoerythrin (PE)/ propidium iodide (PI) filters (10 000 populations, $n = 3$). Any extracellular peptide was released by treatment with trypsin (which also served to detach the cells), while trypan blue was used to quench any extracellular fluorescence,^[10b] thus ensuring that any observed increase in fluorescence was from just the intracellular peptide. a) Flow cytometry histograms gated for live, single cells after incubation with delivery agent as well as propidium iodide (selectively stains the nuclei of necrotic and apoptotic cells^[11]) to exclude dead cells. —: untreated cells, ---: FAM-Ahx-tetraLlp-NH₂,: FAM-Ahx-Glu-Llp-Glu-Glu-NH₂. b) Mean FITC-filtered fluorescence of histograms of the delivery agents versus cell type. Black FAM-Ahx-tetraLlp-NH₂; gray FAM-Ahx-Glu-Llp-Glu-Glu-NH₂. Error bars indicate plus and minus the standard deviation (\pm s.d.)

Table 1: Cell-selective delivery peptides derived from clustering analysis.

	AA ₄	AA ₃	AA ₂	AA ₁
neutrophils	Llp	Llp	Pro	Tyr
monocytes	Glu	Leu	Llp	Llp
lymphocytes	Pro	Lys	Pro	Glu
E14	Glu	Pro	Lys	Llp
K562	Leu	Lys	Lys	Pro
SH-SY5Y	Tyr	Pro	Lys	Leu
HeLa	Pro	Llp	Pro	Glu
HEK293	Leu	Lys	Llp	Lys
ARN8	Pro	Tyr	Glu	Glu

cytometry. In the case of primary cells, the delivery agents were incubated with anti-coagulated whole blood (Figure 4 and Figure S4 in the Supporting Information). Each of the “hit” tetramers resulted in 100% uptake in the targeted cell type (i.e., the entire population was shifted in the histogram, thus illustrating that all cells had internalized the identified peptide), with clear demonstration of selectivity (Figure 4 and Figure S5 in the Supporting Information). Highlighting this result, FAM-Ahx-Leu-Lys-Lys-Pro-NH₂ showed a factor of five higher uptake in K562 cells, FAM-Ahx-Pro-Tyr-Glu-Glu-NH₂ showed a factor of four higher uptake in ARN8 cells, while FAM-Ahx-Glu-Pro-Lys-Llp-NH₂ showed a factor of three higher uptake in E14 cells than in other cell types. Selective uptake between circulating malignant and primary immune cells, which would be of great interest in targeting hematological cancers, was observed with a factor of six higher uptake of FAM-Ahx-Leu-Lys-Lys-Pro-NH₂ in the

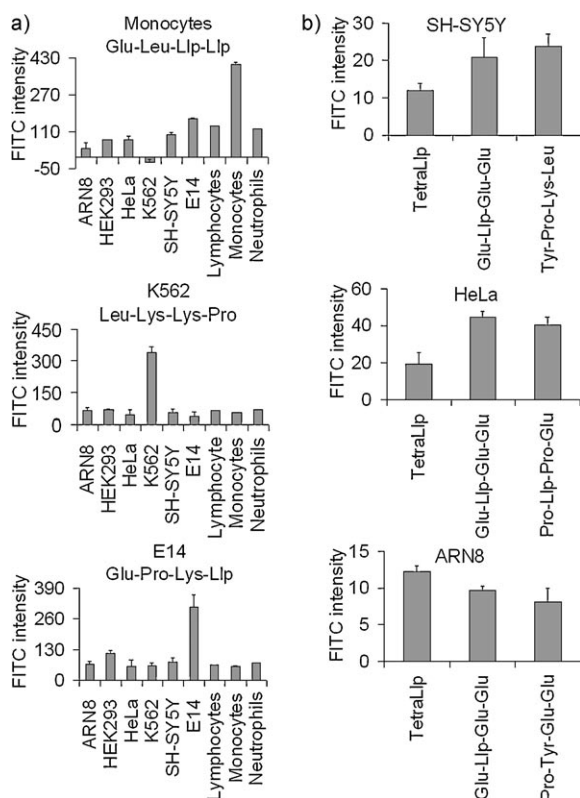


Figure 4. Flow cytometry analysis of the FAM-labeled cell-selective delivery agents (FAM-Ahx-AA₁-AA₂-AA₃-AA₄-NH₂) using FITC and PE-Texas-Red filters (10000 cells, $n = 3$). Conditions as given in Figure 3. Mean FITC-filtered fluorescence of histograms versus a) the cell-selective delivery agents; b) the generic delivery agent compared to the cell-selective delivery agents and positive control (FAM-Ahx-tetraLip-NH₂). Error bars indicate \pm s.d.

myeloid leukemia cell line, K562, than in primary monocytes, which share similar phenotypic characteristics (Figure 4).^[12]

The uptake of some of the hit tetramers was directly compared to the generic consensus peptide FAM-Ahx-Glu-Lip-Glu-Glu-NH₂ and to the positive control FAM-Ahx-tetraLip-NH₂ (Figure 4). The hit tetramers FAM-Ahx-Tyr-Pro-Lys-Leu-NH₂, FAM-Ahx-Pro-Lip-Pro-Glu-NH₂, and FAM-Ahx-Pro-Tyr-Glu-Glu-NH₂ had uptake levels similar to or higher than the positive control and the consensus sequence in the cell lines evaluated, thus illustrating that these hit tetramers are not only selective but also very efficient cell-penetrating peptides.

In order for the peptides to have biological applications, it is vital that they do not exhibit cellular toxicity. Cell viability upon treatment with the hit peptides was assessed with MTT^[13] assays. None of the delivery agents exhibited cytotoxicity in any of the tested cell types at 10 times the concentration of agent used for cell delivery (100 μ M; Figure S7 in the Supporting Information). Additionally, membrane toxicity was assessed using erythrocyte hemolysis assays; no evidence of hemolysis was detected (Figure S7 in the Supporting Information).

To study compartmentalization, the cell-specific delivery peptides were incubated with their respective cell type, and the cells were stained with LysoTracker red and analyzed by

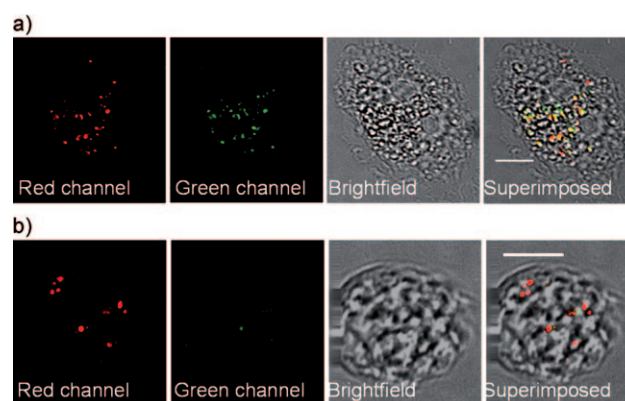


Figure 5. Selected Z-stacks of neutrophil incubated with FAM-Ahx-Glu-Lip-Lip-NH₂ and LysoTracker red (specifically stains the lysosome) at a) 37°C and b) 4°C. Confocal microscopy was performed with FITC and tetramethylrhodamine isothiocyanate (TRITC) filters. Red = LysoTracker red fluorescence, green = FAM-labeled peptide. Yellow/orange color is colocalization of red and green fluorescence. Scale bars = 5 μ m.

confocal microscopy. As a negative control for endocytosis, cells were also incubated with their respective hit tetramer and LysoTracker at 4°C. Superimposed images of the red and green channels revealed colocalization of the peptides (green) and LysoTracker red (yellow), thus illustrating that all the hit tetramers were compartmentalized into the lysosome. In contrast, only the LysoTracker fluorescence in the red channel and no fluorescence in the green channel were observed at 4°C, thus illustrating that internalization of delivery agent was diminished by inhibition of active cellular uptake processes (Figure 5 and Figure S6 in the Supporting Information). Lysosome localization and active transport dependence verifies that all of the delivery agents were taken up by endocytosis.

Microarray analysis of cellular uptake of a 1296-member PNA-encoded peptide library identified an efficient generic delivery agent (Glu-Lip-Glu-Glu) for HeLa, K562, HEK293T, SH-SY5Y, and ARN8 cells; human primary lymphocytes, monocytes, and neutrophils; and embryonic stem cells. This novel anionic delivery agent could in most cases afford better cell penetration than tetraLip.^[10] Furthermore, specific and efficient cell-penetrating homing peptides were identified for these cell types. Highlighting these results, selective uptake between circulating malignant and primary immune cells, which would be of interest in targeting hematological cancers, was observed with factor of six higher uptake of FAM-Ahx-Leu-Lys-Lys-Pro-NH₂ in K562 cells compared to primary monocytes.

Confocal microscopy revealed that the peptide-PNA conjugates were endosomally localized. In combination with the observed selectivity, this finding suggests receptor-mediated endocytosis as the uptake mechanism. Furthermore, the identified peptides showed no toxicity in the tested cell lines or primary cells.

This approach establishes a general strategy for the identification of cell-penetrating peptides as tools or reagents that allow homing to any cell type or tissue of interest, such as

a targeted organ or a tumor. Furthermore, the incorporation of peptoid moieties and PNA encoding makes this screening technique applicable in vivo as well as in vitro, because peptoids and PNAs have long lifetimes in biological environments. In addition, this approach allows identification of ligands for the efficient and cellular targeted delivery of PNA, although we also believe that this technology is not only limited to PNA delivery. Combination of cell-penetrating homing peptides and active pharmacophores might well alleviate the constraints of the properties previously expected of drug molecules, such as small size and cell permeability.^[14]

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- [1] *Drug Delivery—Principles and Applications* (Eds.: B. Wang, T. Siahaan, R. A. Soltero), Wiley, New York, **2005**.
- [2] a) W. Arap, R. Pasqualini, E. Ruoslahti, *Science* **1998**, 279, 377–380; b) R. Haubner, D. Finsinger, H. Kessler, *Angew. Chem.* **1997**, 109, 1440–1456; *Angew. Chem. Int. Ed. Engl.* **1997**, 36, 1374–1389; c) V. Gregorc, A. Santoro, E. Bennicelli, C. J. Punt, G. Citterio, J. N. Timmer-Bonte, F. C. Cappio, A. Lambiase, C. Bordignon, C. M. van Herpen, *Br. J. Cancer* **2009**, 101, 219–224; d) A. Corti, F. Pastorino, F. Curnis, W. Arap, M. Ponzoni, R. Pasqualini, *Med. Res. Rev.* **2011**, DOI: 10.1002/med.20238; e) K. N. Sugahara, T. Teesalu, P. P. Karmali, V. R. Kotamraju, L. Agemy, D. R. Greenwald, E. Ruoslahti, *Science* **2010**, 328, 1031–1035; f) P. Laakkonen, K. Vuorinen, *Integr. Biol.* **2010**, 2, 326–337; g) P. Laakkonen, M. E. Åkerman, H. Biliran, M. Yang, F. Ferrer, T. Karpanen, R. M. Hoffman, E. Ruoslahti, *Proc. Natl. Acad. Sci. USA* **2004**, 101, 9381–9386; h) W. C. Aird, *Circ. Res.* **2007**, 100, 158–173; i) W. C. Aird, *Circ. Res.* **2007**, 100, 174–190.
- [3] D. J. Mitchell, D. T. Kim, L. Steinman, C. G. Fathman, J. B. Rothbard, *J. Pept. Res.* **2000**, 56, 318–325.
- [4] a) A. Astriab-Fisher, D. S. Sergueev, M. Fisher, B. R. Shaw, R. L. Juliano, *Biochem. Pharmacol.* **2000**, 60, 83–90; b) S. Fawell, J. Seery, Y. Daikh, C. Moore, L. L. Chen, B. Pepinsky, J. Barsoum, *Proc. Natl. Acad. Sci. USA* **1994**, 91, 664–668; c) Y. Lim, E. Lee, M. Lee, *Angew. Chem.* **2007**, 119, 3545–3548; *Angew. Chem. Int. Ed.* **2007**, 46, 3475–3478; d) V. P. Torchilin, R. Rammohan, V. Weissig, T. S. Levchenko, *Proc. Natl. Acad. Sci. USA* **2001**, 98, 8786–8791; e) M. Lewin, N. Carlesso, C. H. Tung, X. W. Tang, D. Cory, D. T. Scadden, R. Weissleder, *Nat. Biotechnol.* **2000**, 18, 410–414.
- [5] a) R. F. Service, *Science* **2000**, 288, 28–29; b) S. R. Schwarze, A. Ho, A. Vocero-Akbani, S. F. Dowdy, *Science* **1999**, 285, 1569–1572; c) S. Bacchetti, F. Graham, *Proc. Natl. Acad. Sci. USA* **1977**, 74, 1590–1594; d) *Molecular Biology of the Cell* (Eds.: B. Alberts, A. Johnson, J. Lewis, M. Raff, K. Roberts, P. Walter), Garland Science, New York, **2002**; e) J. E. Murphy, T. Uno, J. D. Hamer, F. E. Cohen, V. Dwarki, R. N. Zuckermann, *Proc. Natl. Acad. Sci. USA* **1998**, 95, 1517–1522.
- [6] M. Mäe, H. Myrberg, S. El-Andaloussi, U. Langel, *Int. J. Pept. Res. Ther.* **2009**, 15, 11–15.
- [7] S. Brenner, R. A. Lerner, *Proc. Natl. Acad. Sci. USA* **1992**, 89, 5381–5383.
- [8] a) N. Svensen, J. J. Díaz-Mochón, M. Bradley, *Tetrahedron Lett.* **2008**, 49, 6498–6500; b) A. Furka, F. Sebestyen, M. Asgedom, G. Dibo, *Int. J. Pept. Protein Res.* **1991**, 37, 487–493.
- [9] a) E. Uhlmann, A. Peyman, G. Breipohl, D. W. Will, *Angew. Chem.* **1998**, 110, 2954–2983; *Angew. Chem. Int. Ed.* **1998**, 37, 2796–2823; b) S. W. Jones, R. Christison, K. Bundell, C. J. Voyce, S. M. V. Brockbank, P. Newham, M. A. Lindsay, *Br. J. Pharmacol.* **2005**, 145, 1093–1102.
- [10] a) A. Unciti-Broceta, F. Diezmann, C. Y. Ou-Yang, M. A. Fara, M. Bradley, *Bioorg. Med. Chem.* **2009**, 17, 959–966; b) V. L. Mosiman, B. K. Patterson, L. Canterero, C. L. Goolsby, *Cytometry* **1997**, 30, 151–156.
- [11] H. Lecoeur, *Exp. Cell Res.* **2002**, 277, 1–14.
- [12] a) E. Klein, H. Ben-Bassat, H. Neumann, P. Ralph, J. Zeuthen, A. Polliack, F. Vánky, *Int. J. Cancer* **1976**, 18, 421–431; b) B. B. Luzzio, C. B. Luzzio, E. G. Bamberger, A. S. Feliu, *Proc. Soc. Exp. Biol. Med.* **1981**, 166, 546–550.
- [13] T. Mosmann, *J. Immunol. Methods* **1983**, 65, 55–63.
- [14] J. P. Richard, K. Melikov, E. Vives, C. Ramos, B. Verbeure, M. J. Gait, L. V. Chernomordik, M. Lebleu, **2003**, 278, 585–590.
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